

Whole-genome sequencing of a *Janthinobacterium sp.* isolated from the Patagonian Desert

Nicole T. Cavanaugh,¹ Girish Kumar,² Alicyn Reverdy Pearson,¹ Julia Colbert,¹ Carlos Riquelme,³ André O. Hudson,² Yunrong Chai,¹ Veronica Godoy-Carter¹

AUTHOR AFFILIATIONS See affiliation list on p. 2.

ABSTRACT *Janthinobacterium* is a genus of Gram-negative environmental bacteria that survive extreme conditions by forming biofilms and producing pigments. *Janthinobacterium sp.* LS2A, an extremophile isolated from soil in the Chilean Patagonia, contains seven known biosynthetic gene clusters, including the purple pigment violacein, which may aid in its survival in harsh environments.

KEYWORDS environmental microbiology, soil microbiology, genomics, extremophiles

Janthinobacterium is a genus of rod-shaped Gram-negative bacteria that is widespread in cold environments (1–3). Some produce a purple pigment called violacein (4). Violacein is of high biotechnological significance due to its known antiviral, antibacterial, antimycotic, antitumor, algicidal, and antioxidant activities (5–8). The genus also contains extremophiles that are tolerant to temperatures 2%–28°C and UV exposure (9–11). *Janthinobacterium sp.* also form biofilms, which help them survive these harsh conditions (4).

The goal of this project was to study extremophiles in Patagonia and the mechanisms that help them survive in extreme environments. Soil from Laguna Sarmiento (51.0636 S; 72.9264 W) was collected in July 2017. Bacteria were isolated after plating 100 µL of a soil/water slurry on Reasoner's 2A (R2A) agar plates, followed by incubation at 28°C for 48–72 hours. Several bacterial colonies emerged; a single purple colony was picked and assigned the name “LS2A.” To test biofilm formation, LS2A was grown at 30°C to OD600 = 1.0 in R2A broth, and 2 µL of the culture was spotted on a dry R2A plate and incubated for 96 hours before imaging (Fig. 1A). As a first approximation to identify closely related species, we performed 16S rRNA gene sequencing on the V3/4 variable regions using the following primers: 5'-CCTACGGGNGGCWGCAG-3' and 5'-GACTACHVGGGTATCTAATCC-3'. The isolate was identified as *Janthinobacterium sp.* using the National Society for Biotechnology Information (NCBI) BLASTN version 2.15.0 (12, 13). The sequences of the V3/4 16S rRNA region, along with a variety of 16S sequences from *Janthinobacterium*

TABLE 1 Predicted BGCs in LS2A^a

Region	Type	Start	End	Most Similar To	Similarity Score
1.1	RiPP-like	364,750	372,021	Burkholderic acid	Low (<10%)
1.2	Indole	533,201	556,218	Violacein	100%
1.3	RiPP-like	830,949	841,878		
1.4	Acyl amino acids	936,131	1,031,826	O-antigen	14%
1.5	Terpene	1,087,968	1,109,729		
3.1	RiPP-like	254,757	266,376		
7.1	Arylpolyene	1	32,414	APE Ec	36%

^aA summary of biosynthetic gene clusters (BGCs) in *Janthinobacterium sp.* LS2A identified by antiSMASH.

Editor Simon Roux, DOE Joint Genome Institute, Berkeley, California, USA

Address correspondence to Veronica Godoy-Carter, v.godyocarter@northeastern.edu.

The authors declare no conflict of interest.

See the funding table on p. 3.

Received 17 June 2024

Accepted 8 September 2024

Published 9 October 2024

Copyright © 2024 Cavanaugh et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

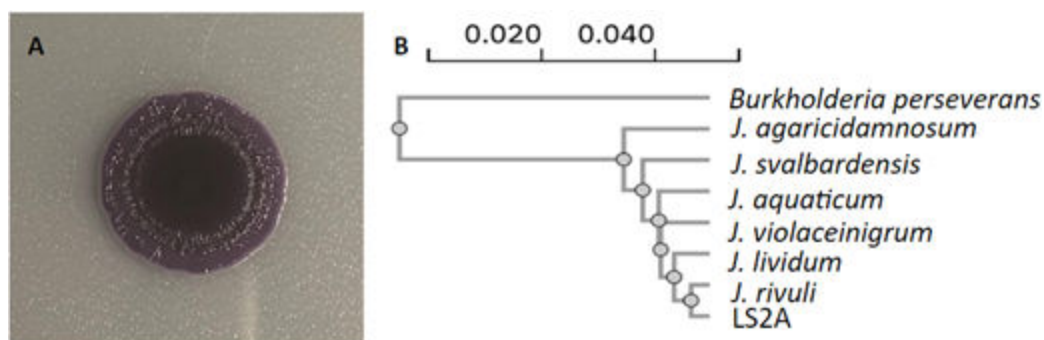


FIG 1 *Janthinobacterium* sp. LS2A forms biofilms, and its sequence demonstrates it belongs to the genus. (A) Biofilm grown from *Janthinobacterium* sp. LS2A. (B) Phylogenetic tree. A phylogenetic tree relating the LS2A V3/4 16S rRNA regions to other *Janthinobacterium* sp. in the RefSeq database. From top to bottom: *Burkholderia perseverans* NR_179094.1, *J. agaricidamnosum* NR_114134.1, *J. svalbardensis* NR_132608.1, *J. aquaticum* NR_170539.1, *J. violaceinigrum* NR_170541.1, *J. lividum* NR_026365.1, *J. rivuli* NR_170540.1, and *Janthinobacterium* sp. LS2A.

sp. from the RefSeq database, were used to create a multiple sequence alignment and guide tree using ClustalOmega version 1.2.4 (Fig. 1B) (14).

Genomic DNA was isolated from a 2-mL R2A liquid culture grown at 30°C overnight using the Wizard Genomic DNA Purification Kit (Promega, USA) following the manufacturer's instructions for Gram-negative bacteria. DNA libraries were prepared using the Nextera XT library preparation kit (Illumina) and sequenced using the Illumina MiSeq System at the Genomics Lab, Rochester Institute of Technology. Raw paired-end reads were trimmed using Trimmomatic Galaxy version 0.38.1 (15). The trimmed reads were assembled *de novo* using Unicycler Galaxy version 0.5.0+galaxy1 (16). Quality analysis was performed using Quast Galaxy version 5.2.0+galaxy1 (17) and FastQC Galaxy version 0.74 + galaxy0 (18). All programs listed above were run using default parameters unless stated otherwise. The assembly consists of 4,575,745 reads totaling 667.6 million base pairs (Mbp). These reads assembled into 29 contigs with a total length of 6,251,303 base pairs (bp). The estimated genome coverage is 105 x with 98.45% completeness. The N50 is 540,701 bp, and the GC content is 62.69%. Analysis by the Prokaryotic Genome Analysis Pipeline version 6.6 revealed that the LS2A genome contained 5,664 total genes, 5,548 protein-coding sequences, three rRNAs operons, and 77 tRNAs (19–21).

A summary of the biosynthetic gene clusters and secondary metabolites predicted by antibiotics and secondary metabolite analytics shell (antiSMASH) version 7.1.0 is outlined in Table 1 (22). antiSMASH detected seven gene clusters that encode unique metabolites, including violacein.

ACKNOWLEDGMENTS

We would like to thank the National Science Foundation (MCB1651732) and the NU Global Experience Office for their support. Thank you to the RIT Genomics Lab. V.G.-C. was supported by an HHMI Inclusive Excellence grant and the Northeastern Global Experiential Office. J.C. was supported by NU Undergraduate Research and Fellowships Office PEAK Awards. N.T.C. was supported by the NSF Graduate Research Fellowship Program (1938052). A.R.P. was supported by the Northeastern University Provost Dissertation Completion Fellowship.

AUTHOR AFFILIATIONS

¹Department of Biology, Northeastern University College of Science, Boston, Massachusetts, USA

²Rochester Institute of Technology College of Science, Thomas H. Gosnell School of Life Sciences, Rochester, New York, USA

³Departamento de Biotecnología, Universidad de Antofagasta, Antofagasta, Chile

AUTHOR ORCID*s*

Nicole T. Cavanaugh  <http://orcid.org/0009-0001-6602-5062>

André O. Hudson  <http://orcid.org/0000-0001-5690-4322>

Yunrong Chai  <http://orcid.org/0000-0002-5903-5529>

Veronica Godoy-Carter  <http://orcid.org/0000-0002-1254-9282>

FUNDING

Funder	Grant(s)	Author(s)
National Science Foundation (NSF)	MCB1651732	Yunrong Chai
Howard Hughes Medical Institute (HHMI)	52008706	Veronica Godoy-Carter

AUTHOR CONTRIBUTIONS

Nicole T. Cavanaugh, Data curation, Formal analysis, Investigation, Methodology, Project administration, Validation, Writing – original draft, Writing – review and editing | Girish Kumar, Data curation, Formal analysis, Investigation, Methodology, Validation | Alicyn Reverdy Pearson, Data curation, Investigation, Methodology | Julia Colbert, Investigation, Methodology | Carlos Riquelme, Supervision, Writing – review and editing | André O. Hudson, Funding acquisition, Supervision, Writing – review and editing | Yunrong Chai, Conceptualization, Project administration, Supervision, Writing – review and editing | Veronica Godoy-Carter, Conceptualization, Funding acquisition, Investigation, Methodology, Project administration, Writing – review and editing

DATA AVAILABILITY

The WGS projects for *Janthinobacterium* sp. LS2A have been deposited in GenBank under BioProject number [PRJNA1071647](https://ncbi.nlm.nih.gov/bioproject/PRJNA1071647), BioSample number [SAMN39706722](https://ncbi.nlm.nih.gov/biosample/SAMN39706722), and SRA number [SRR27839980](https://ncbi.nlm.nih.gov/sra/SRR27839980). Results from PGAP analysis can be found using accession number [JAZHPB000000](https://ncbi.nlm.nih.gov/assembly/JAZHPB000000). The following sequences were used to construct Fig. 1 and were retrieved from GenBank: [NR_179094.1](https://ncbi.nlm.nih.gov/nuccore/NR_179094.1), [NR_114134.1](https://ncbi.nlm.nih.gov/nuccore/NR_114134.1), [NR_132608.1](https://ncbi.nlm.nih.gov/nuccore/NR_132608.1), [NR_170539.1](https://ncbi.nlm.nih.gov/nuccore/NR_170539.1), [NR_170541.1](https://ncbi.nlm.nih.gov/nuccore/NR_170541.1), [NR_026365.1](https://ncbi.nlm.nih.gov/nuccore/NR_026365.1), and [NR_170540.1](https://ncbi.nlm.nih.gov/nuccore/NR_170540.1).

The NCBI BLASTN webserver was used to identify the LS2A 16s rRNA gene sequence. All programs found on the Galaxy platform were run using Galaxy version 23.1 and can be found at usegalaxy.org. Clustal Omega can be found on the [EMBL European Bioinformatics Institute](https://www.ebi.ac.uk/EMBL-EBI/EMBL-EuropeanBioinformaticsInstitute) webpage.

REFERENCES

- Chernogor L, Bakhvalova K, Belikova A, Belikov S. 2022. Isolation and properties of the bacterial strain *Janthinobacterium* sp. SLB01. *Microorganisms* 10:1071. <https://doi.org/10.3390/microorganisms10051071>
- Gong X, Skrivergaard S, Korsgaard BS, Schreiber L, Marshall IPG, Finster K, Schramm A. 2017. High quality draft genome sequence of *Janthinobacterium psychrotolerans* sp. nov., isolated from a frozen freshwater pond. *Stand Genomic Sci* 12:8. <https://doi.org/10.1186/s40793-017-0230-x>
- Schloss PD, Allen HK, Klimowicz AK, Mlot C, Gross JA, Savengsuksa S, McEllin J, Clardy J, Ruess RW, Handelsman J. 2010. Psychrotrophic strain of *Janthinobacterium lividum* from a cold alaskan soil produces prodigiosin. *DNA Cell Biol* 29:533–541. <https://doi.org/10.1089/dna.2010.1020>
- Pantanello F, Berlutti F, Passariello C, Sarli S, Morea C, Schippa S. 2007. Violacein and biofilm production in *Janthinobacterium lividum*. *J Appl Microbiol* 102:992–999. <https://doi.org/10.1111/j.1365-2672.2006.03155.x>
- Andrighetti-Fröhner CR, Antonio RV, Creczynski-Pasa TB, Barardi CRM, Simões CMO. 2003. Cytotoxicity and potential antiviral evaluation of violacein produced by *Chromobacterium violaceum*. *Mem Inst Oswaldo Cruz* 98:843–848. <https://doi.org/10.1590/s0074-02762003000600023>
- Konzen M, De Marco D, Cordova CAS, Vieira TO, Antônio RV, Creczynski-Pasa TB. 2006. Antioxidant properties of violacein: possible relation on its biological function. *Bioorg Med Chem* 14:8307–8313. <https://doi.org/10.1016/j.bmc.2006.09.013>
- Durán N, Justo GZ, Ferreira CV, Melo PS, Cordi L, Martins D. 2007. Violacein: properties and biological activities. *Biotechnol Appl Biochem* 48:127–133. <https://doi.org/10.1042/BA20070115>
- Cai G, Yang X, Yu X, Zheng W, Cai R, Wang H. 2024. The novel application of violacein produced by a marine *Duganella* strain as a promising agent for controlling *Heterosigma akashiwo* bloom: algicidal mechanism, fermentation optimization and agent formulation. *J Hazard Mater* 466:133548. <https://doi.org/10.1016/j.jhazmat.2024.133548>
- Yang M, Lu D, Qin B, Liu Q, Zhao Y, Liu H, Ma J. 2018. Highly efficient nitrogen removal of a coldness-resistant and low nutrient needed

- bacterium, *Janthinobacterium* sp. M-11. *Bioresour Technol* 256:366–373. <https://doi.org/10.1016/j.biortech.2018.02.049>
10. Mukhia S, Kumar A, Kumari P, Kumar R. 2022. Psychrotrophic plant beneficial bacteria from the glacial ecosystem of sikkim himalaya: genomic evidence for the cold adaptation and plant growth promotion. *Microbiol Res* 260:127049. <https://doi.org/10.1016/j.micres.2022.127049>
 11. Mojib N, Farhoomand A, Andersen DT, Bej AK. 2013. UV and cold tolerance of a pigment-producing antarctic *Janthinobacterium* sp. Ant5-2. *Extremophiles* 17:367–378. <https://doi.org/10.1007/s00792-013-0525-9>
 12. Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res* 25:3389–3402. <https://doi.org/10.1093/nar/25.17.3389>
 13. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <https://doi.org/10.1186/1471-2105-10-421>
 14. Madeira F, Pearce M, Tivey ARN, Basutkar P, Lee J, Edbali O, Madhusoodanan N, Kolesnikov A, Lopez R. 2022. Search and sequence analysis tools services from EMBL-EBI in 2022. *Nucleic Acids Res* 50:W276–W279. <https://doi.org/10.1093/nar/gkac240>
 15. Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>
 16. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>
 17. Mikheenko A, Prjibelski A, Saveliev V, Antipov D, Gurevich A. 2018. Versatile genome assembly evaluation with QUAST-LG. *Bioinformatics* 34:i142–i150. <https://doi.org/10.1093/bioinformatics/bty266>
 18. Babraham bioinformatics—FastQC a quality control tool for high throughput sequence data. Available from: <https://www.bioinformatics.babraham.ac.uk/projects/fastqc/>. Retrieved 2 Feb 2024.
 19. Haft DH, DiCuccio M, Badretdin A, Brover V, Chetvernin V, O'Neill K, Li W, Chitsaz F, Derbyshire MK, Gonzales NR, Gwadz M, Lu F, Marchler GH, Song JS, Thanki N, Yamashita RA, Zheng C, Thibaud-Nissen F, Geer LY, Marchler-Bauer A, Pruitt KD. 2018. RefSeq: an update on prokaryotic genome annotation and curation. *Nucleic Acids Res* 46:D851–D860. <https://doi.org/10.1093/nar/gkx1068>
 20. Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvernin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the prokaryotic genome annotation pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>
 21. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>
 22. Blin K, Shaw S, Augustijn HE, Reitz ZL, Biermann F, Alanjary M, Fetter A, Terlouw BR, Metcalf WW, Helfrich EJN, van Wezel GP, Medema MH, Weber T. 2023. antiSMASH 7.0: new and improved predictions for detection, regulation, chemical structures and visualisation. *Nucleic Acids Res* 51:W46–W50. <https://doi.org/10.1093/nar/gkad344>